# EASTERN US CONTINENTAL SHELF CARBON BUDGET

### Integrating Models, Data Assimilation, and Analysis

BY THE USECOS TEAM

### US EAST COAST CONTINENTAL SHELF (USECoS) PROJECT TEAM.

Eileen Hofmann (hofmann@ccpo.odu.edu) is Professor, Department of Ocean, Earth and Atmospheric Sciences, Old Dominion University, Norfolk, VA, USA. Jean-Noël Druon is NRC Postdoctoral Scientist, NASA Goddard Space Flight Center, Greenbelt, MD, USA, and is currently at IFREMER, Center at Brest, DYNECO-Benthos Laboratory, Plouzané, France. Katja Fennel is Assistant Professor, Department of Oceanography, Dalhousie University, Halifax, Nova Scotia, Canada. Marjorie Friedrichs is Research Assistant Professor, Virginia Institute of Marine Science, Gloucester Point, VA, USA. Dale Haidvogel is Professor, Institute of Marine and Coastal Sciences, Rutgers University, New Brunswick, NJ, USA. Cindy Lee is Professor, Marine Sciences Research Center, Stony Brook University, Stony Brook, NY, USA. Antonio Mannino is Oceanographer, NASA Goddard Space Flight Center, Greenbelt, MD, USA. Charles McClain is SeaWiFS Project Scientist, NASA Goddard Space Flight Center, Greenbelt, MD, USA. Raymond Najjar is Associate Professor, Department of Meteorology, Pennsylvania State University, University Park, PA, USA. John Siewert is Research Associate, Department of Meteorology, Pennsylvania State University, University Park, PA, USA. John O'Reilly is Research Ecologist, NOAA/NMFS Narragansett Laboratory, Narragansett, RI, USA. David Pollard is Senior Research Associate, Earth and Environmental Systems Institute, Pennsylvania State University, State College, PA, USA. Michael Previdi is Postdoctoral Research Associate, Institute of Marine and Coastal Sciences, Rutgers University, New Brunswick, NJ, USA, and is currently at Lamont-Doherty Earth Observatory, Columbia University, Palisades, NY, USA. Sybil Seitzinger is Professor, Institute of Marine and Coastal Sciences, Rutgers University, New Brunswick, NJ, USA. Sergio Signorini is Senior Scientist, Science Applications International Corporation, Beltsville, MD, USA. John Wilkin is Associate Professor, Institute of Marine and Coastal Sciences, Rutgers University, New Brunswick, NJ, USA.

THE PAST TWO DECADES have seen the development of large multidisciplinary oceanographic programs that focus on understanding carbon cycling processes in coastal and oceanic environments. Synthesis and modeling activities typically followed toward the ends of these programs (e.g., Joint Global Ocean Flux Study), usually long after the field experiments had been planned and carried out. A lesson from these programs was articulated in subsequent community planning reports (e.g., the Ocean Carbon Transport, Exchanges and Transformations Report [OCTET; http://www.msrc.sunysb.edu/octet/ Workshop\_Report.htm] and Ocean Carbon and Climate Change Report [OCCC; http://www.carboncyclescience. gov/documents/occc is 2004.pdf]): future ocean carbon cycle research programs should promote close collaborations among scientists with expertise in measurement, data analysis, and numerical modeling at every stage of development-formative stages of hypothesis building, planning and execution of field programs, data analysis, numerical

modeling, and synthesis.

The advantages of this collaborative approach for numerical model development are clear: data are collected with algorithm development in mind, and quantitative models are based on the best current understanding. But there are distinct advantages for observational and experimental programs as well. Collaborations with numerical modelers allow analysts to see data in a more holistic context, and thus to understand better what other parameters should be measured and modeled and what are the inherent limitations and uncertainties in the modeling approaches. That the collaboration of modelers and analysts results in better model construction, with justifiable assumptions, and in more appropriate data selection has been recognized for some time (e.g., Walsh, 1972); it is re-emphasized by recent discussions of approaches for development of marine biogeochemical and ecosystem models (e.g., Doney, 1999; Doney et al., 2001; Flynn, 2005; Anderson, 2005). For an analytical measurement where no standards exist, it may be only through sensitivity studies with numerical models that the analyst can predict what the values of certain parameters are likely to be. Modeling and data-intensive programs often have the same goals, but they bring different tools to bear. The box models and statistical techniques common to data analyses are part of the quantitative view of data. But using all the methods available (e.g., empirical and deterministic approaches) will ensure greater progress toward the common goal of understanding large ecological systems.

We combined the expertise of modelers and empiricists in a collaborative project as part of the NASA Earth

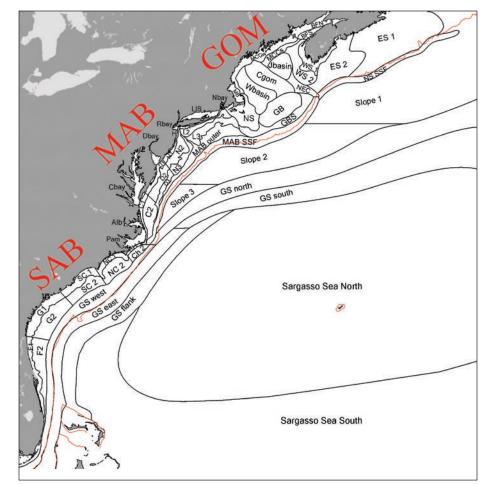


Figure 1. Map of the US East Coast showing the Middle Atlantic Bight (MAB), the South Atlantic Bight (SAB), the Gulf of Maine (GOM), the 58 subregions used for evaluation of model-derived distributions, and the 500-m isobath (red line). GB = Georges Bank, GS = Gulf Stream, SSF = shelf slope front, NEC = Northeast Channel, Wbasin = Wilkinson Basin, Jbasin = Jordan Basin, Cgom = central Gulf of Maine.

Interdisciplinary Science initiative. The US East Coast Continental Shelf (USECoS) project was initiated in 2004 with the overall goal of developing carbon budgets for the Mid-Atlantic Bight (MAB) and South Atlantic Bight (SAB) along the eastern US coast (Figure 1). We addressed this goal through a series of specific research questions that were designed to understand carbon inputs and fates in the two regions, dominant food-web pathways for carbon cycling, and similarities/differences in carbon cycling in the two continental shelf systems. The nature of the research questions required a team approach that included expertise in areas of remote sensing, oceanographic data analysis, numerical models including data assimilation, carbon cycling, and knowledge of the physics and biogeochemistry of the MAB and SAB. As a result, the team assembled for the USECoS project consists of a diverse group of science investigators with varying degrees of experience in crossing disciplinary boundaries. This mixture of expertise and the interactions that resulted has proven to be as important in successfully addressing the project goal as any infrastructure (e.g., computers), data sets, and numerical model codes that we used. However, the fruitful scientific collaborations have come with a steep learning curve.

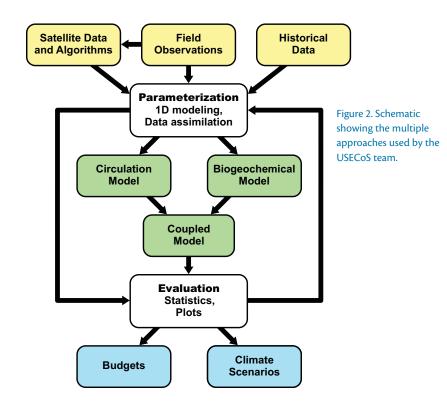
Integrating results from different disciplines and expertise included using measurements from satellites, field studies, historical data, and one-dimensional data assimilative modeling. Simplified mathematical descriptions (parameterizations) were developed to capture the essential features of each disciplinary model; these parameterizations were then implemented in the circulation and biogeochemical models used in this study (Figure 2). Satellite-derived data products were evaluated with field and historical data to ensure their accuracy; results of parameterization studies were incorporated into the circulation and biogeochemical models; simulation results (models run separately and coupled) were evaluated using a suite of approaches that included escalating statistical evaluations; and results of the evaluation phase were used to revise parameterizations. This iterative process of model improvement and evaluation (Figure 2) continued until simulations were deemed sufficiently realistic, and thus ready to provide the bases for development of nutrient and carbon budgets and to serve as baselines for climate-related simulations.

The primary objective of this paper is to give insight into how collaborations between analysts and modelers strengthened a program that is yielding results that likely would not have been achieved without them. We illustrate these collaborations with examples of how model simulations and processes were evaluated using comparisons with historical in situ and satellite-derived data sets, quantitative statistical estimates of model skill, and data assimilation. The accompanying text boxes provide details of the Northeast North America (NENA) circulation model (Box 1), the biogeochemical model (Box 2), some of the specific methods used for quantitative skill assessment (Box 3), the dissolved organic matter (DOM) model (Box 4), and the satellite dissolved organic carbon (DOC) algorithm development program (Box 5) within the USECoS program.

This paper also demonstrates how the ongoing USECoS program is enhancing our understanding of carbon cycling processes on the MAB and SAB continental shelves. This research is particularly germane to NASA's carbon cycle research focus and coastal research initiative and the US Climate Change Research Program, all of which support the goals of the North American Carbon Program (Wofsy and Harriss, 2002). We highlight primary approaches used as well as some of the challenges and results that have come from interactions among our team of investigators. Addressing the global scale and interdisciplinary nature of the science questions now facing Earth scientists requires integrated teams of investigators to answer them. Thus, the lessons we have learned provide insights and a way forward for future programs.

### MAB AND SAB CIRCULATION AND PRODUCTIVITY PATTERNS

The continental shelf of the eastern United States is a relatively well-studied region of the ocean; there is abundant historical data for a first-order physical and biogeochemical characterization.



Carbon cycling in the MAB and SAB continental shelves and upper slope has been studied for 30 years in a number of programs sponsored by the Department of Energy (DOE, 1970s and 1980s). Much of the DOE-sponsored work in the SAB is summarized in Atkinson et al. (1985), and some of the earliest studies using the Coastal Zone Color Scanner (CZCS) were conducted in collaboration with the DOE SAB program (Yoder et al., 1987; McClain et al., 1988). In the MAB, the Shelf Edge Exchange Processes (SEEP) experiments I and II (1983-1989) and the Ocean Margins Program (OMP) experiment (1994-1996) provided insight into biogeochemical processes, with major findings reported in special issues of Continental Shelf Research (1988, 8[5-7]) and Deep-Sea Research Part II (1994, 41[2-3]; 2002, 49[20]). Yoder et al. (2001) used the entire 7.5-year CZCS data set to examine phytoplankton variability of the MAB and the SAB and showed noticeable interannual variability during 1978-1986.

From these past studies, we know that the circulation dynamics and the productivity and chlorophyll fields in the MAB and SAB differ significantly. In the SAB, the Gulf Stream flows along the outer edge of the shelf break (e.g., Lee and Atkinson, 1983), producing upwelling with subsurface bottom intrusions and frontal eddies that have a strong effect on nutrient and plankton production (Yoder, 1985). The episodic forcing of the SAB by Gulf Stream-induced upwelling results in biological production that occurs in short-lived events (McClain et al., 1984) rather than the more traditional spring/fall blooms that are observed in the MAB. In summer, when SAB shelf waters are stratified and

the Gulf Stream tends to be nearer the shelf break, the intrusions extend across the entire shelf and produce subsurface blooms that are not discerned in oceancolor imagery. In winter, when shelf waters are well mixed, satellite-derived ocean-color distributions from the SAB show the episodic nature of the chlorophyll production in this region and phytoplankton blooms adjacent to the MAB coast in some years. The differences between these two regions provide a strong basis for comparative studies between a continental shelf region that is strongly affected by oceanic forcing (SAB) and one in which buoyancy and wind forcing are more dominant (MAB).

USECoS study participants seek to

...collaborations between analysts and modelers strengthened a program that is yielding results that likely would not have been achieved without them.

suggest that it occurs in multiple sites along the outer SAB shelf.

In contrast to the SAB, the MAB has an outer shelf front and a slope sea that separate the shelf proper from the Gulf Stream. The influence of the Gulf Stream on the MAB is through warm-core eddies that move southward along the shelf break (Evans et al., 1986). The shelf circulation in both systems is influenced by estuarine and riverine inputs and wind. In the MAB, the coastal flow is to the south, with offshore flow at Cape Hatteras, where much of the flow is entrained into the Gulf Stream front. Cross-shelf exchange occurs along the entire shelf edge through meandering of the shelf-break front (Lozier and Gawarkiewicz, 2001), and is at times modulated by warm-core-ring interactions (Ryan et al., 2001). Ocean-color distributions from the MAB show an annual April-May spring bloom (Yoder et al., 2001), as well as extensive summer understand how carbon is introduced into the eastern US continental shelf environment, how it is transformed and transported while resident on the shelf, and what is its ultimate fate. Our approach to these questions is to use (1) remote sensing data, especially ocean-color imagery from the Sea-viewing Wide Field-of-View Sensor (SeaWiFS) and the Moderate Resolution Imaging Spectroradiometer (MODIS), (2) a synthesis of in situ measurements, (3) a coupled ocean biogeochemistrycarbon-circulation model configured for the MAB and SAB, and (4) dataassimilation studies.

### MODEL-DATA FUSION

Model-data fusion embraces a number of approaches for integrating discrete observations into a modeling framework, ranging from simple model-data comparisons to more formal data assimilation methodologies such as

35

### BOX 1: CIRCULATION MODEL DESCRIPTION

Realistic simulation of the circulation on the MAB and SAB continental shelves is fundamental to realizing the objectives of the USECoS program; thus, considerable effort has been directed at achieving acceptable simulations. The circulation model used is the Regional Ocean Modeling System (ROMS) (Shchepetkin and McWilliams, 2005; Haidvogel et al., in press; http://myroms. org) configured for the same Northeast North America (NENA) spatial domain of Fennel et al. (2006). The model has 10-km horizontal resolution, 30 vertical levels, and is embedded within the HYbrid Coordinate Ocean Model (HyCOM, http://hycom.org) North Atlantic data assimilative model (Chassignet et al., 2007). The HyCOM open boundary transports are augmented by barotropic tides from a global analysis (Egbert and Erofeeva, 2002). Coastal freshwater inputs have annual mean values from a watershed analysis (Seitzinger et al., 2005) modulated by average monthly variability observed at US Geological Survey-gauged rivers. Air-sea fluxes are calculated using bulk formulae (Fairall et al., 2003) applied to daily reanalysis of air temperature, pressure, humidity, and winds. All tracer advection is by the positive definite multidimensional positive definite advection transport algorithm (MPDATA) scheme, which is important for accurate representation of biogeochemical model constituents.

The embedding procedure imposes external, remotely forced mesoscale and seasonal variability, but achieving realistic mean circulation in shelf waters proved critical to correct biases in the temperature and salinity provided by the HyCOM North Atlantic model. A simple correction procedure—supplanting the HyCOM temporal mean temperature and salinity with values from the Hydrobase climatology (Lozier et al., 1995)—was devised that substantially improved the simulation of buoyancy-driven southwestward mean flow throughout the Gulf of Maine and MAB. Improvements to the properties of slope water adjacent to the SAB (which enters the NENA domain from the Intra-American Seas) were also noted.

The simulations exhibit well-recognized features of the local and remotely forced circulation: low salinity on the MAB inner shelf, the tidal mixing front and residual circulation around Georges Bank, Gulf Stream intrusions in the SAB, and interactions of Gulf Stream warm rings with the New England slope. Comparisons of the modeled circulation and tracer fields to observations show progress over the simulations of Fennel et al. (2006), due principally to the introduction of tides and unbiased open boundary data. Features that are the focus of ongoing study are exaggerated upwelling of anomalously cold water in the SAB, intermittent overshoot of the Gulf Stream at Cape Hatteras, and a weak Slope Sea gyre. Model development is investigating whether higher spatial resolution is required to improve these aspects of the simulation. constrained parameter optimization. We have had success with a number of these approaches because of the extensive and continual collaboration between our observational (in situ and satellite) and modeling science investigators.

Quantitative model skill assessment (Figure 2) is a significant activity that engages all components of the USECoS team. Our collective experiences indicate that this activity is frequently not a major component of multidisciplinary team research. Simulations from the NENA model have been evaluated by comparison with in situ and satellitederived data using a suite of statistical approaches of escalating rigor, including comparisons of spatial distributions, means, variance, two-dimensional histograms, and other skill assessment methods, such as Target (Joliff et al., in press) and Taylor (Taylor, 2001) diagrams, which reveal spatial and/or seasonal timing/phase relationships. Overall spatial distributions of climatological means from the model should match those from in situ and satellite data with little bias; the model should also capture the dynamic range over seasonal time scales, as well as regional differences in the timing of minima and maxima such as spring and fall phytoplankton blooms. This diversity of model skill assessment methods has helped us identify seasons and regions where model improvements are required. In addition, a one-dimensional data assimilative model has provided the basis for quantitative assessment of model processes, which furthers the development of a model with improved skill. Each approach is described below, and their results provide the bases for evaluation of model processes.

### Model Evaluation Through Historical Data Comparisons

We focused our historical data-mining efforts on temperature, salinity, and dissolved oxygen measurements in the 2005 World Ocean Database (WOD, Boyer et al., 2006), selecting only those data that have been interpolated to standard levels and that passed all WOD quality-control procedures. For our study region (Figure 1, excluding the Sargasso Sea), there are about 460,000 temperature profiles, 110,000 salinity profiles, and 20,000 oxygen profiles. The median year of the temperature station distribution is 1968; 90% of the profiles were made between 1946 and 1994; similar characteristics apply to the salinity and oxygen data.

Using these data, we developed monthly mean climatologies of mixed layer depth (MLD), salinity, and dissolved oxygen anomaly ( $\Delta O_2$ , departure from saturation), which were used to evaluate equivalent distributions constructed from our circulation and biogeochemical model (Figure 3). For example, our circulation model (Box 1) captures a number of the observed patterns in MLD such as: (1) large parts of the shelf and Georges Bank are well mixed to the bottom in March; (2) in the MAB, MLD increases away from the shelf, sometimes exceeding 250 m, but then reaches a minimum of about 50 m within the extension of the Gulf Stream northeast of Cape Hatteras; and (3) in September, the mixed layer is shallower and tends to deepen away from the continent, except for Georges Bank, where tides mix to the bottom. The annual mean salinity distribution is also well simulated by the circulation

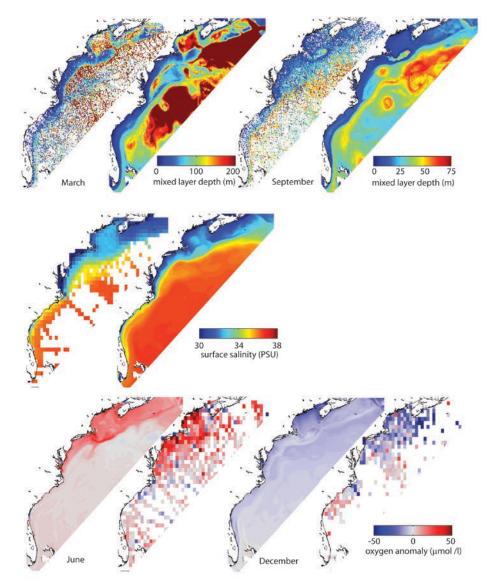


Figure 3. Climatological and model-derived distributions of mixed-layer depth (upper panel), annual mean salinity (middle panel), and dissolved oxygen anomaly (lower panel). Mixed-layer depth (MLD) was computed using a criterion of 0.5°C with respect to the surface. Monthly MLD was binned to a 0.1° grid; monthly salinity and oxygen anomaly were binned to a 0.5° grid. Annual mean salinity was computed when a given grid box contained measurements for more than nine calendar months. White indicates no data.

model (Figure 3), showing the dominant pattern of increasing surface salinity with distance from shore, a large salinity gradient located near the shelf break, low salinity on the Scotian shelf, and high salinity in the Sargasso Sea.

Historical data analyses are also integral to the evaluation of the marine biogeochemical model (Box 2). Dissolved oxygen, particularly its departure from saturation, has long been used as a tracer for the cycling of organic carbon. We computed the oxygen anomaly using the WOD temperature, salinity, and dissolved oxygen data, with a formulation for the saturation concentration (Garcia

37

### BOX 2: BIOGEOCHEMICAL MODEL STRUCTURE

We simulate ecosystem processes in the MAB and SAB with a modified version of the Fasham et al. (1990) model that is incorporated into ROMS (Fennel et al., 2006). In our ROMS implementation, we distinguish the two inorganic nitrogen species, nitrate and ammonium; include chlorophyll as a prognostic variable in addition to phytoplankton biomass; distinguish two size classes of detritus to allow for different settling rates; and include explicit DOM dynamics described in detail in Box 4.

It is important to note that none of the biological models available in ROMS explicitly represents diagenetic processes at present. However, the inclusion or at least the parameterization of diagenetic processes is important for coastal applications because a major fraction of nutrient remineralization occurs in the sediment. In our application of the Fasham-type model to the East Coast continental shelves (western North Atlantic) we use a relatively simple representation of benthic remineralization processes where organic matter settling out of the bottommost grid box results in a corresponding influx of inorganic nutrients at the sediment/water interface (Fennel et al., 2006). This formulation conserves mass by assuming immediate equilibrium between particle deposition and influx of dissolved constituents from the sediment. Soetaert et al. (2000) showed that this intermediate complexity approach captures most of the dynamics inherent in benthic-pelagic coupling when compared to coupling with a diagenetic submodel, but is computationally much more efficient. This approach also allows for the straightforward inclusion of processes such as sediment denitrification (Fennel et al., 2006) using the relationship between sediment oxygen consumption and denitrification derived by Seitzinger and Giblin (1996).

We also included inorganic carbon and oxygen dynamics in our biogeochemical model for the MAB and SAB. Aside from physical transport (i.e., advection and mixing), the local concentration of dissolved inorganic carbon (DIC) and oxygen is affected by gas exchange with the atmosphere at the sea surface and by sources and sinks caused by biological processes such as the photosynthetic synthesis of organic matter or its metabolic or microbial oxidation. We describe the sources and sinks due to biological processes based on stoichiometric ratios and parameterize gas exchange as suggested by Wanninkhof (1992). The gas exchange of oxygen depends on the temperature-and-salinity-dependent oxygen solubility (Garcia and Gordon, 1992) and the piston velocity. For the air-sea gas exchange of carbon dioxide, the situation is more complicated because gas exchange does not directly depend on the concentration of DIC but rather on the small fraction of DIC that is present as carbon dioxide (carbon dioxide does not just dissolve in seawater, but rather reacts with water to form carbonic acid, which subsequently dissociates into bicarbonate and carbonate; the sum of all three makes up DIC). Only the small fraction of DIC that is present in the form of carbon dioxide determines the partial pressure, pCO<sub>2</sub>, that enters the parameterization of gas exchange. Calculating this fraction (and thus pCO<sub>2</sub>) requires knowledge of DIC, the local alkalinity, temperature and salinity, and the iterative solution of a set of nonlinear equilibrium equations (Zeebe and Wolf-Gladrow, 2001).

and Gordon, 1992), and compared it with the similar quantity obtained from the biogeochemical model (Figure 3). The model captures the overall pattern of the surface ocean oxygen anomaly, reproducing the observed supersaturation in early summer and undersaturation in early winter, with particularly large seasonal ranges inshore of the shelf break north of Cape Hatteras. The annual cycle in oxygen anomaly reflects the annual cycle in surface heat flux, net community production, and vertical mixing. Because the circulation model captures the annual cycle in MLD and salinity very well, model-data differences are most likely due to biogeochemical processes not yet represented.

### Model Evaluation Through Satellite Data Comparisons Satellite-Model Comparisons

We are also using a wide range of satellite-derived distributions (Figure 4, Table 1) to quantify and understand regional, seasonal, interannual, and climate-related variability of phytoplankton biomass and organic carbon production within the USECoS study area and to evaluate the performance of similar products from the simulations with the coupled circulation-biogeochemical model. We have developed several new and simple metrics that characterize the natural cycles of major annual phytoplankton biomass and carbon production events. One example is the index of "month of maximum satellite chlorophyll concentration" (Figure 4D), which was computed from a nine-year monthly SeaWiFS climatology data set. This case reveals that the fall phytoplankton bloom (September and October) in the northern Gulf of Maine is a more significant

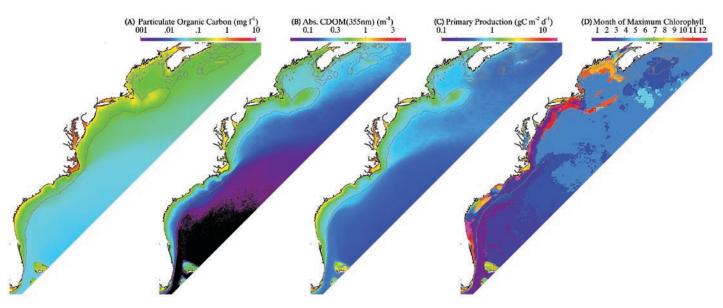


Figure 4. The nine-year mean distribution of (A) particulate organic carbon (POC), (B) absorption by colored dissolved organic matter at 355 nm (a<sub>cdom</sub>), (C) primary production (Vertically Generalized Productivity Model 2a, VGPM2a), and (D) month of maximum concentration chlorophyll *a* constructed from SeaWiFS measurements made from 1997 to 2007.

event in the annual cycle than the spring bloom there. Peak chlorophyll occurs during January in the inner MAB shelf but during November in the mid-shelf, and during April along the outer MAB shelf and adjacent Slope Sea. On Georges Bank, the annual chlorophyll peak occurs in April, except along the northern and southern flanks of the Bank, adjacent to the tidal mixing fronts, where the peak occurs in October-November. There is also a surprising degree of spatial heterogeneity in the timing of the annual peak in the SAB. Satellite data and simple metrics such as these reveal the relevant underlying biological oceanographic scales operating on the continental shelf and reveal important differences in processes among the SAB, MAB, Georges Bank, and the Gulf of Maine regions.

Comparisons of satellite-derived fields with equivalent fields from NENA show that the model captures the north-south gradient in sea surface temperature (SST,

Measurements	Sources
Sea Surface Temperature (SST, °C)	4 km nighttime composite from: • AVHRR Pathfinder 1985–present • MODIS Terra 2000– • MODIS Aqua 2002– • GOES 2001–
Chlorophyll <i>a</i> (mg m <sup>-3</sup> )	SeaWiFS and MODIS Aqua
Photosynthetically Active Radiation (PAR, E $m^{-2} d^{-1}$ )	SeaWiFS
Cloud Cover/Probability	SeaWiFS
Particulate Organic Carbon (POC, mg L <sup>-1</sup> )	D. Clark Algorithm (uses SeaWiFS/MODIS ocean-color bands ~ 443 nm, ~ 490 nm, ~ 550 nm)
Primary Production (g C m <sup>-2</sup> d <sup>-1</sup> ) Euphotic Depth (1% surface par) Euphotic Chlorophyll (mg m <sup>-2</sup> )	SeaWiFS data and the VGPM2a model VGPM2a model VGPM2a model
Absorption Coefficient of CDOM (m <sup>-1</sup> ) a <sub>CDOM</sub> (355 nm) (m <sup>-1</sup> ) Dissolved Organic Carbon (μmol L <sup>-1</sup> )	SeaWiFS and MODIS (see Box 5) SeaWiFS and MODIS (see Box 5)

Table 1. Satellite data and derived products used for analyses and model evaluation.

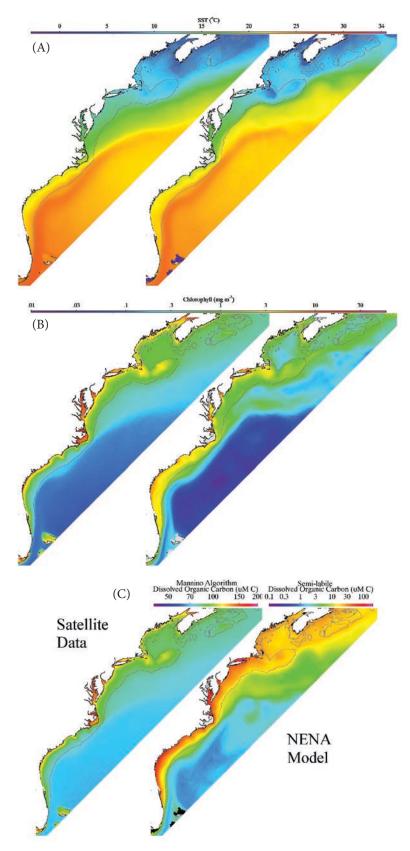


Figure 5. Comparisons of satellite-derived (left panels) and simulated (right panels) distributions obtained from the Northeast North America (NENA) model of annual mean (A) sea-surface temperature (SST), (B) chlorophyll, and (C) dissolved organic carbon (DOC) in the surface water and simulated semilabile DOC.

Figure 5A) and the general north-south gradient in chlorophyll distribution, but underestimates concentrations in the SAB except in the mid-shelf (Figure 5B); it also captures the spatial pattern in DOC concentration (Figure 5C). These comparisons provide a first order evaluation of model skill and highlight areas where model improvements are needed.

### A Cautionary Note on Estimating Productivity from Satellite Data

The ability to estimate primary productivity (PP) from space enables the determination of phytoplankton carbon production for the world ocean with unprecedented temporal and spatial resolutions. Accurate measurement of surface chlorophyll a, SST, and photosynthetically active radiation (PAR) are key elements of nearly all satellite oceancolor-based primary productivity algorithms (Carr et al., 2006). In shelf water north of Cape Hatteras, the broad-scale seasonal and spatial patterns from the VGPM2a satellite-productivity model (a variation of the Vertically Generalized Productivity Model; Behrenfeld and Falkowski, 1997) are consistent with results based on in situ measurements (<sup>14</sup>C uptake) made during earlier surveys (O'Reilly et al., 1987). However, the SAB continental shelf poses a unique challenge for satellite-based PP estimates because the episodic summer subsurface intrusions of nutrient-rich Gulf Stream waters onto the shelf significantly enhance biomass and carbon production below the depths "visible" to passive satellite ocean-color sensors, such as SeaWiFS and MODIS.

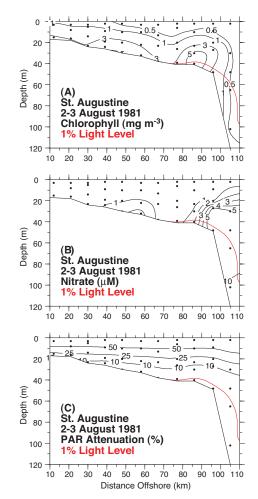
Vertically integrated chlorophyll *a* and PP within two weeks after a large bottom intrusion event on the middle

40

shelf in July 1981 reached 75 mg m<sup>-2</sup> and 3 to 4 g C m<sup>-2</sup> d<sup>-1</sup>, respectively (Yoder et al., 1985). At the peak of the bloom, 80% of the water column PP occurred below the mixed layer, and new PP (NO<sub>3</sub>-supported) exceeded 90% of the total. Most of this production occurred below the mixed layer, as well as below the penetration depth (the inverse of the diffuse attenuation coefficient) of oceancolor sensors, making it impossible to use satellite ocean-color algorithms to observe the effects of these sub-surface blooms on SAB phytoplankton production (Signorini et al., 2005).

As an example, cross sections of chlorophyll *a*, nitrate, and light penetration from a transect off St. Augustine, FL, (Figure 6A) show pronounced intrusion effects. Below 20 m, where Gulf Stream intrusions enhance the nitrate concentration (Figure 6B), and sufficient light (1 to 10% of surface PAR, Figure 6C) is available for photosynthesis, biomass (Figure 6A) and carbon production (not shown) increase significantly toward the bottom. The subsurface bloom intensity varies from station to station in response to the magnitude of the nutrient enrichment originating from the intrusion.

The mean chlorophyll *a* concentration estimated from the CZCS-derived chlorophyll *a* summer composite for 1981 (Figure 6D) was 0.32 mg m<sup>-3</sup> versus an in situ estimate of 0.45 mg m<sup>-3</sup> for the St. Augustine section. Near the shelf edge, where in situ near bottom chlorophyll *a* concentrations were largest (> 5 mg m<sup>-3</sup>), the satellite coverage was good, but provided no hint of the subsurface bloom. These results illustrate the need for more than one approach for estimating rates of carbon production by phytoplankton in continental shelf waters



and highlight the importance of combining satellite-derived estimates with those from mechanistic models that can extend measurements to deeper waters.

## Model Evaluation Through Target and Taylor Diagrams

To evaluate the model, we are making standard side-by-side comparisons of model-data contour plots (Figures 3 and 5) and time series of selected simulated and observed quantities at specific depths and locations. In addition,

(D) Chlorophyll a Concentration C19811721981263.L3m\_SNSU\_CHLO\_4

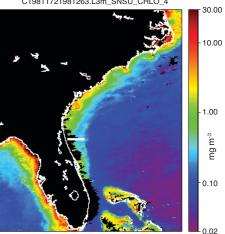


Figure 6. Cross-shelf sections of: (A) chlorophyll a, (B) nitrate, and (C) light penetration (photosynthetically active radiation [PAR] in percent of surface intensity) constructed from data collected during August 2-3, 1981, along a transect offshore of St. Augustine, FL (30°N). The light penetration was calculated using a PAR attenuation coefficient developed for the ecosystem model, which is a function of chlorophyll a concentration and salinity. Note the high chlorophyll *a* concentration on the entire shelf below 10 m, which resulted from nutrient inputs from a subsurface Gulf Stream intrusion, and significant (> 10%) light penetration to the ocean floor. A Coastal Zone Color Scanner (CZCS) summer composite for 1981 (D) shows that ocean-color retrievals miss the high chlorophyll *a* that is associated with the subsurface bottom intrusions (thick white line shows transect location). Note that there are no satellite data in the transect region near the coast (black regions) due to sensor amplifier ringing off the bright coastline.

however, we are applying more quantitative measures, such as Target and Taylor diagrams (Box 3), to assess model skill.

Target diagrams (Joliff et al., in press) are used to visualize the relative magnitudes of the two components of root mean square differences (RMSD) (i.e., the misfits of the means and of the variability). Target diagrams for SST and surface chlorophyll (Figure 7) show that these components of the total RMSD are typically of equal magnitude, though higher for chlorophyll than SST. Within

### BOX 3: TARGET AND TAYLOR DIAGRAMS

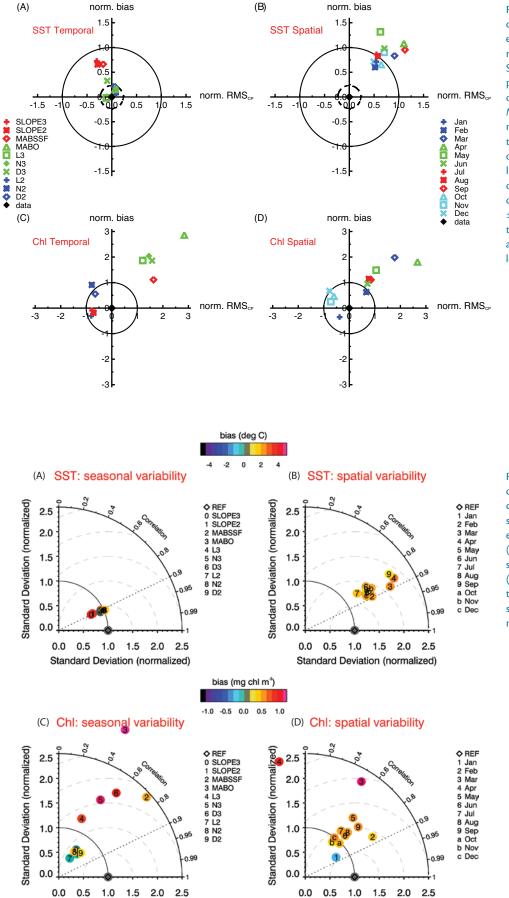
Target (Joliff et al., in press) and Taylor diagrams (Taylor, 2001) quantitatively compare and visualize statistics associated with multidimensional time series (e.g., SST or surface chlorophyll in Figure 7). Typically used to compare models and data, they might also contrast observations derived from different sources. These diagrams are based on computations of the square root of mean square differences (RMSD) between model and data. The RMSD is composed of two components, the bias representing the difference between the means of the two fields, and the centered-pattern RMSD (RMSD<sub>CP</sub>) representing differences in variability.

The Target diagram (Joliff et al., in press) exploits the fact that RMSD squared is equal to the RMSD CP squared and the bias squared. Thus, if these two statistics are plotted on the x- and y-axes, respectively, the distance from the origin is equal to the total RMSD. Though RMSD<sub>CP</sub> is a positive quantity, on Target diagrams RMSD<sub>CP</sub> is given the sign of the difference between the standard deviation of the model and observed time series. Thus, negative values of RMSD<sub>CP</sub> indicate that the model underestimates the observed variability, while positive values indicate that the model variability is an overestimate. If these quantities are normalized by the standard deviation of the observations, it is possible to compare model-data fit of monthly averages for multiple subregions (Figure 7A, C), or compare modeled and observed spatial patterns for given months (Figure 7B, D). In addition, when normalized in this manner, symbols falling within the total RMSD = 1 circle indicate the model provides a better estimate of productivity than the mean of the observations. An inner circle (Figure 7A, B) representing the uncertainty associated with the observations can also be included in Target diagrams; when points lie within this inner-circle (the bull's-eye), model and data are by definition statistically indistinguishable from each other.

The RMSD<sub>CP</sub> can be further decomposed into contributions from phase differences between the two time series (or cross correlation) and differences in their standard deviations (Oke et al., 2002). The Taylor diagram (Taylor, 2001) quantitatively compares these three statistics for a given distribution (e.g., SST or chlorophyll; Figure 8). The normalized standard deviation of the simulated distribution is the radial distance from the origin, the correlation of the two distributions is the angle from the x-axis, and the normalized centered-pattern root-mean-square difference is then the distance between the data symbol at location [1,0] on the x-axis and each model symbol (indicated by the dashed lines, in the same units as the coordinates). Bias information is thus not inherently depicted on a Taylor diagram, but is included here by using colored symbols (Figure 8). As in the Target diagrams, Taylor diagrams can illustrate temporal (Figure 8A, C) or spatial (Figure 8B, D) variability in model-data ft.

each of our 58 subregions (Figure 1), the model reproduces the seasonal variability of SST very well, in many cases to within the uncertainty of the observations. In terms of the spatial variability of SST (Figure 7B), the model overestimates both the mean and the variability of the observations-a result stemming from the simulated Gulf Stream being located closer to the shelf than the observations indicate (Figure 5). Similar trends appear for the spatial variability of surface chlorophyll (Figure 7D), with the model overestimating the mean and variance of the SeaWiFS chlorophyll data during the spring bloom, yet underestimating the variance during the fall.

Taylor diagrams (Figure 8) indicate that within each subregion, the time series of monthly (2004) average simulated SST are well correlated with satellite data (with a correlation coefficient of  $\sim 0.9$ ). The model time series have nearly the same variance as the observations, except in the slope waters where the model tends to underestimate seasonal variability of SST (Figure 8A). In terms of the spatial distributions, the model overestimates the differences in SST between the various subregions, especially in March, April, and September (Figure 8B). Not surprisingly, the model has less skill in reproducing the seasonal variability of chlorophyll, as compared with SST, especially in the outer MAB and the MAB shelf-slope front where the model significantly overestimates the observed variability. In terms of the spatial distributions, the model accurately reproduces the observed differences between the subregions in summer and late fall, but overestimates the variability in spring. In April, the model results are inversely



Standard Deviation (normalized)

Standard Deviation (normalized)

Figure 7. Target diagrams showing the bias, centered-pattern root mean square difference (RMSD), and total RMSD (Box 3) for monthly (2004) model vs. satellite-derived SST (upper panels) and chlorophyll (lower panels). (A) and (C): model-data misfit of time-series within 10 subregions of the MAB. (B) and (D): monthly model-data misfit in terms of spatial distributions within the same 10 subregions. Circles denote lines of constant normalized total RMSD. Solid line: model-data misfit = standard deviation of data. Dashed line in (A) and (B): modeldata misfit = error in data, assumed to be ± 0.5°C. Thus, model estimates falling within the inner circle (bull's-eye) are indistinguishable in terms of skill. Region definitions in legend refer to regions defined in Figure 1.

Figure 8. Taylor diagrams (Box 3) showing quantitative assessment of model skill when compared to satellite data (from 2004) for sea surface temperature (SST) (upper panels) and surface chlorophyll (lower panels). (A) and (C): model-data misfit of timeseries within 10 subregions of the MAB. (B) and (D): monthly model-data misfit in terms of spatial distributions within the same 10 subregions. Bias is illustrated via model symbol colors.

correlated with the observations, a result that has helped to focus the direction of our current efforts to improve the model.

### Model Refinement Through One-Dimensional Data Assimilation

Another approach taken to incorporate discrete observations into the NENA framework and to quantitatively assess model skill is the variational adjoint method (Friedrichs et al., 2006, 2007). Due to our incomplete knowledge, biogeochemical models are often by necessity highly empirical, have many nonmechanistic formulations, and include numerous parameters that are difficult to measure with current oceanographic instrumentation. Data assimilation techniques such as the variational adjoint method (Hofmann and

(A)

Friedrichs, 2001) provide an approach for objectively estimating the best-fit set of model parameters and their associated uncertainties (Fennel et al., 2001). These methods can be used to compute sensitivities and correlations between parameters and assess predictive abilities of a given model (Friedrichs et al., 2006), and are thus a crucial component of successful marine biogeochemical modeling studies.

We are currently making use of an existing one-dimensional data assimilative ecosystem-modeling framework that has been recently developed to quantitatively compare the performance of 12 models characterized by varying levels of ecosystem complexity (Friedrichs et al., 2007). When used in conjunction with the three-dimensional NENA model, which provides the horizontal advection terms, vertical velocity, MLD, PAR, and temperature, this framework closely reproduced the three-dimensional fields and yielded optimal values of maximum phytoplankton growth rate, remineralization rates, C:chl ratios, and other key parameters (Figure 9). Because this framework includes the flexibility of assimilating remotely sensed ocean-color and/or in situ data simultaneously from multiple sites (Friedrichs et al., 2007), it is possible to estimate parameter values that provide the best fit to multiple regions simultaneously. Data can also be assimilated from numerous individual locations (Friedrichs et al., 2006), revealing whether certain optimal parameter values and their associated uncertainties vary in space and/or time.

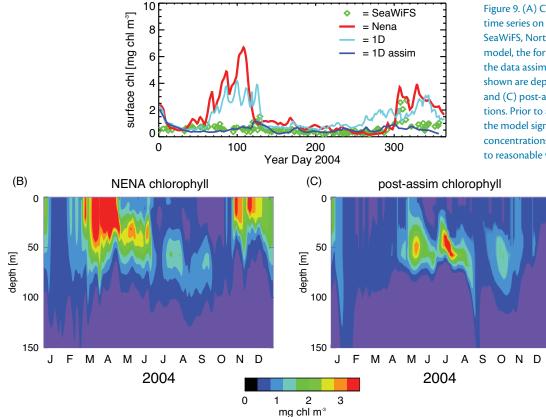


Figure 9. (A) Comparison of surface chlorophyll time series on the outer New Jersey shelf from: SeaWiFS, Northeast North America (NENA) model, the forward one-dimensional model, and the data assimilative one-dimensional model. Also shown are depth-time contour plots of (B) NENA and (C) post-assimilation chlorophyll concentrations. Prior to assimilation of ocean-color data, the model significantly overestimated chlorophyll concentrations in the spring. These were reduced to reasonable values by adjusting the maximum

> growth rate, the initial slope of the photosynthesis versus irradiance curve, and the mortality rate of phytoplankton; however, these parameter changes did not improve the fit for the short-lived fall bloom (year day 306–316), which may reveal missing complexity in the ecosystem dynamics.

### EVALUATION OF MODEL PROCESSES

In addition to using historical data sets to evaluate distributions of various concentrations predicted by our models, we also evaluated process mechanisms. For example, it is known that DOM represents the largest pool of organic carbon on the MAB shelf and may thus play a significant role in carbon cycling and transport. Furthermore, under nutrient-depleted conditions, nitrogen and carbon primary production are partially decoupled; the DOM produced is carbon-rich and thus may represent a significant source of organic carbon (Williams, 1995), which can be exported to the open ocean (Bauer and Druffel, 1998; Vlahos et al., 2002). To investigate this aspect of shelf carbon cycling, we included semilabile dissolved organic carbon and nitrogen (semilabile DOC and DON) in our biogeochemical model (Box 4). The inclusion of explicit DOM dynamics allows investigation of its role in biological production and carbon cycling in the MAB and SAB in conjunction with physical transport, which is difficult to do with just observations (Box 5).

Analysis of our model predictions allowed us to assess the relative importance of DOM production and transport through advective and eddy-diffusive mechanisms. The simulated annual horizontal divergence of semilabile DOC shows areas of production and export (negative values) on the continental shelf and slope and areas of import (positive values) in the open ocean (Figure 10A). Most shelf areas can export about 1 mol C m<sup>-2</sup> yr<sup>-1</sup> through the seasonal production of marine semilabile DOC to the open ocean.

### BOX 4: INCLUSION OF DOM DYNAMICS IN THE BIOGEOCHEMICAL MODEL

Because a large portion of organic carbon is stored in dissolved organic matter (DOM), the transport of DOM may be an important pathway for carbon. As a first step toward addressing this question, we added two semilabile DOM components (dissolved inorganic nitrogen [DON] and dissolved inorganic carbon [DOC]) to the biogeochemical model. Note that the model does not include the biologically inert "refractory" DOM fraction that dominates DOM in deep waters and is thought, except in areas influenced by rivers (Druffel et al., 1992), to act as a conservative tracer and be relatively uniform with depth (Hansell and Carlson, 1998; Carlson, 2002). The source and sink terms of DOM are phytoplankton exudation, "sloppy feeding" of zooplankton, POM solubilization, and DOM remineralization. The sources and sinks of DOM are thus directly related to primary production, grazing, and detritus pool concentration.

Exudation of semilabile DOC by phytoplankton includes two processes: nutrient-based and carbon-excess-based release. The nutrient-based release reflects the exudation of semilabile DOC and DON by healthy phytoplankton. This term is proportional to primary production. The carbon-excess-based release represents carbohydrate over-production by nutrient-stressed cells. This process is responsible for the mucilage events that are often observed during summer in eutrophic coastal areas. The carbon excess uptake can be seen as an "overflow" of photosynthesis under nutrient limitation. We described this process as the difference between nutrient-saturated (and light-limited) and nutrient-limited (and light-limited) primary production (Andersen and Williams, 1998; Ianson and Allen, 2002).

As a result, in nutrient-depleted conditions, the nitrogen primary production and carbon primary production are partially decoupled and carbonrich DOM is produced; C-to-N ratios for DOM range between 10 and 25 (Hopkinson and Vallino, 2005; Søndergaard et al., 2000; Biddanda and Benner, 1997; Benner et al., 1992). Values vary from 9.95 for fresh material (Hopkinson and Vallino, 2005) to 19–25 for high molecular weight DOM (i.e., mainly the semilabile fraction [Biddanda and Benner, 1997]). The accumulation and subsequent transport of carbon-rich DOM may thus present an efficient mechanism for export of organic carbon from productive shelf systems to the open ocean.

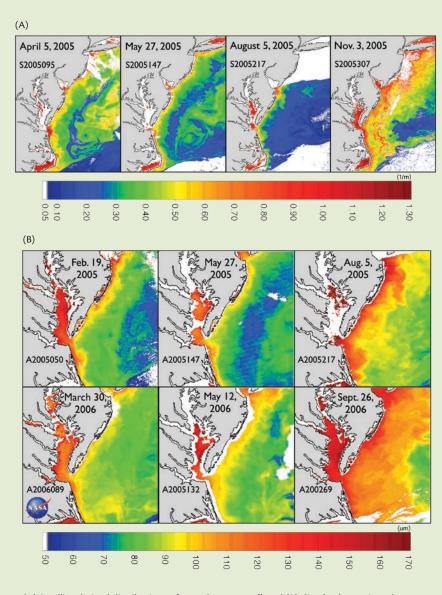
Thomas et al. (2002) suggest that the US northeastern continental shelf is an important site for particulate organic matter (POM) burial. In order to compare the magnitudes of POM burial and the horizontal export of DOM to the open ocean, we added to the biogeochemical model a parameterization mimicking POM resuspension and burial. Resuspension rate of the POM flux reaching the seabed was specified as a function of the bottom friction velocity. The fraction of resuspended POM is thus largely dependent on the local nearbottom current velocity associated with the general circulation, tidal currents on the continental shelf, and wind-driven events in shallow waters, and it further couples the physical and biogeochemical models. We used carbon and nitrogen burial to simulate the accumulation of material in the sediment, assuming that the burial efficiency of the particulate organic carbon (POC) is proportional to the vertical flux of POC reaching the seabed (Henrichs and Reeburgh, 1987). We estimated the burial efficiency for particulate organic nitrogen (PON) using a C to N ratio of buried organic matter of 9.3; values of 9-10 have been found for shelf and estuarine surface sediments and slightly lower values in deeper waters (Gelinas et al., 2001). The model gave burial rates of POC in the sediments (Figure 10A) that agreed well with estimates by Thomas et al. (2002) of 0.1–0.2 mol C m<sup>-2</sup> yr<sup>-1</sup> in the slope off Cape Cod, 0.5-1.0 in the Mid-Atlantic Bight, and 0.02-1.7 in the slope off Cape Hatteras. The main gradients of model organic carbon burial extended from 2 mol C m<sup>-2</sup> yr<sup>-1</sup> in shallow water (inner shelf) to 0.2 mol C m<sup>-2</sup> yr<sup>-1</sup> on the outer shelf.

### BOX 5: DEVELOPMENT OF EMPIRICAL ALGORITHMS FOR CDOM AND DOC

As part of a limited USECoS field program focused on the Chesapeake Bay region, samples were collected at multiple depths for measurement of pigments, dissolved organic carbon (DOC), particulate organic carbon (POC), absorption of chromophoric dissolved organic matter (CDOM), and particles. These samples have provided data sets critical for development of satellite-derived DOC and POC algorithms and for evaluation of the biogeochemical model results (Figure 5).

In order to develop empirical algorithms for CDOM and DOC, we collected field measurements to correlate  $a_{CDOM}$  (CDOM absorption coefficient) to in situ radiometry (reflectance band ratios) and then correlated DOC to reflectance band ratios through the  $\mathbf{a}_{_{\text{CDOM}}}$  to DOC relationships. Results from our current fieldwork in the continental margin of the southern MAB and the mouth of Chesapeake Bay demonstrate that we can retrieve  $\mathbf{a}_{_{\text{CDOM}}}$  from SeaWiFS and MODIS observations through empirical relationships similar to those described by other researchers (D'Sa and Miller, 2003; Johannessen et al., 2003). Our a CDOM algorithm takes an exponential decay form with the remote-sensing reflectance band ratios (488/551 nm for MODIS and 490/555 nm for SeaWiFS) plotted on the ordinate and a CDOM on the abscissa; author Mannino, M.E. Russ, and S.B. Hooker are preparing a paper on satellite-derived distributions of DOC and CDOM in the US Middle Atlantic Bight. Because CDOM contributes to light absorption across the visible spectrum, several band ratio solutions are possible to avoid the atmospheric correction problems associated with the 412-nm, ocean-color satellite band in coastal waters (e.g., negative water-leaving radiances). Furthermore, comparisons of CDOM absorption at other relevant wavelengths (e.g., 443 nm) are possible, for example, with the Garver-Siegel-Maritorena inversion model (GSM-01; Maritorena et al., 2002). Uncertainties for a CDOM derived from MODIS-Aqua are on average 20–25% and < 10% for DOC (Mannino manuscript noted above). Seasonal variability in CDOM absorption and DOC is quite evident along the continental margin, with the estuarine plumes and near-shore regions as most dynamic (Box 5 figure). Our results show that at least two seasonal algorithms (fall-winter-spring and summer) are required to retrieve DOC from MODIS and SeaWiFS due to seasonal variability in the CDOM-to-DOC relationship caused by the accumulation of primarily nonchromophoric DOC from net ecosystem production (NEP) and the concomitant loss of CDOM through sunlight-induced photooxidation between late spring and early fall.

Satellite ocean-color data and field measurements are helping us to evaluate results from our biogeochemical model. For example, the increase in DOC distributions observed from ocean-color satellite data can be compared with model results on the seasonal accumulation of semilabile DOC from net ecosystem productivity (Figures 4 and Box 5 figure, part B). The satellite ocean-color data are also helping us to evaluate how well the model represents the inputs and fate of DOC to the continental shelf from estuarine and riverine systems.



(A) Satellite-derived distributions of  $a_{CDOM}$  (355 nm, m<sup>-1</sup>) and (B) dissolved organic carbon (DOC) ( $\mu$ M C). From spring to summer,  $a_{CDOM}$  decreases due to photooxidation and possibly from reduced inputs of terrigenous DOM as a result of reduced river discharge. From summer to fall, storm events will vertically mix the water column and introduce chromophoric dissolved organic matter (CDOM) from depth into surface waters. Much higher DOC values are observed in summer compared to early spring due to ecosystem productivity that promotes the accumulation of semilabile DOC. Our estimate of the DOC reservoir within the 10–100-m isobaths for the continental shelf region shown in Figure 1 is on the order of 1.2 Tg C. Source data for the images shown are from NASA's SeaWiFS and MODIS-Aqua sensors.

Comparison of simulated POC burial and horizontal flux of semilabile DOC (Figure 10) shows that POC is efficiently buried in the inner and mid shelf while the mid- and outer-shelf export seasonally produced DOC to the open ocean at comparable rates. This simple parameterization gives us the opportunity to assess the importance of these processes quantitatively in our coupled circulation-biogeochemical model. Our results suggest that the inclusion of a more comprehensive sediment transport and transformation model (Warner et al., in press) should be an important future focus in model refinement.

### CONCLUDING REMARKS

The USECoS project represents a major effort to simultaneously synthesize and integrate diverse data sets, field measurements, models, and modeling approaches. We expect that the type of approach taken here will result in more insight than would be possible if each component of the program moved forward independently. The primary significance of this project is in providing a strong quantitative basis for the development of future observational and modeling studies of carbon budgets of continental shelf systems. A strong aspect of the USECoS project is that the integration of modeling and extensive physical, chemical, and biological data sets provides an opportunity for modeling efforts and data analyses to inform one another from the outset.

The extensive collaboration between the in situ and satellite data analysts and modelers has improved our parameterizations and formulations for both the circulation and biogeochemical models, and it has identified areas where

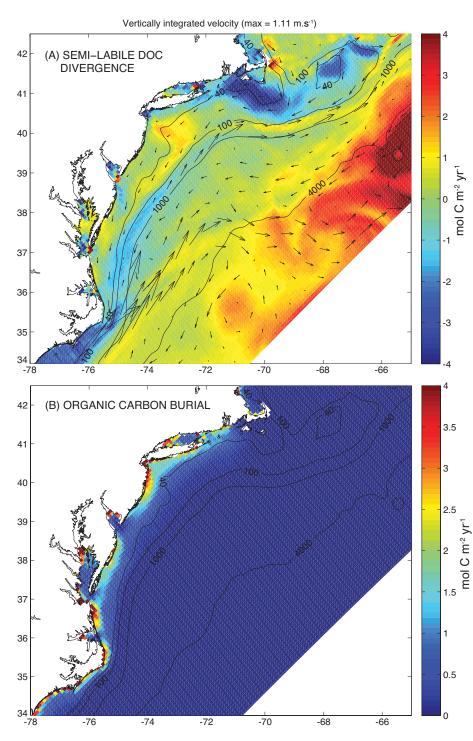


Figure 10. (A) Simulated semilabile dissolved organic carbon (DOC) net horizontal flux for 2005 (mol C  $m^2$  yr<sup>1</sup>). Negative values correspond to areas of production and export, and positive values to areas of import. (B) Simulated particulate organic carbon (POC) burial in the sediments for 2005 (mol C  $m^2$  yr<sup>1</sup>).

improvements in satellite algorithms may be needed (e.g., primary production). The example of estimating primary production in SAB shelf waters resulting from bottom intrusions highlights the need for coupled numerical circulationbiogeochemical models capable of providing accurate estimates of primary production in continental shelf waters, especially since the SAB is not the only continental shelf system where much of the primary production is at depths greater than can be seen by ocean-color satellites (e.g., Prézelin et al., 2004). However, model development should be undertaken in conjunction with in situ measurements of primary production made with current technology to provide rigorous evaluations of the model-derived estimates, similar to the approaches used in this study. This task may not be easy to accomplish under the current funding limitations, but coordinated model-data efforts are needed if reducing uncertainties is a goal.

Empirical observations of process measurements such as primary production of POC and DOC, benthic primary production, remineralization rates of DOM, solubilization and remineralization of particles, burial efficiency in shallow and deep waters, and grazing-related release of DOM are critical to improving biogeochemical models as model complexity increases. Model evaluation is usually focused around measurements of concentration and biomass (cf. Figures 4, 5, 7, 8), but model development needs measurements of rates and processes. For example, few rate measurements were available to constrain the processes included in the DOM component of NENA. The available data sets allow autotrophic processes in the NENA

region to be fairly well constrained, but there are few direct measurements of heterotrophic processes, such as those involved in decomposition of organic matter. The coastal ocean is one place where heterotrophic processes are large enough to be measured. Without such measurements, models such as NENA will continue to include ill-constrained parameters, which results in a trialand-error approach for developing model parameterizations and processes, thereby limiting model skill. However, the measurements and biogeochemical model development are not independent and should progress together to ensure realization of the full benefit of each.

A primary conclusion from the approach taken in this study is that a well-coordinated interdisciplinary team with skills in field measurements, remote sensing, and modeling focused on a single coupled circulation-biogeochemical model is an effective means of addressing important and complex issues, such as carbon cycling in marine ecosystems. The focus on a single model forces the team to resolve issues and reconcile differences of opinion (i.e., a disciplined approach, rather than simply going in different directions, as can happen with a focus on more than one model).

A research team composed of members from multiple institutions, like the USECoS team, does at times hinder progress. However, a team that is dispersed may be unavoidable in achieving the desired balance of expertise. Maintaining progress requires a commitment from each team member to interactive collaborations, and there is a trade-off between having a critical mass and having a team that is too large to manage. A large team can lead to development of smaller groups focused on specific research problems that do not foster collaborative interactions. Many factors must come together to make a successful program (NRC, 2005); for the USECoS program, a focus on one model and common goals has allowed the overall effort to be more than the sum of the individual components.

### ACKNOWLEDGEMENTS

The USECoS project is supported by the National Aeronautics and Space Administration Interdisciplinary Science Program. ₪

#### REFERENCES

- Anderson, T.R. 2005. Plankton functional type modelling: Running before we can walk? *Journal of Plankton Research* 27:1,073–1,081.
- Anderson, T.R., and P.J. le B. Williams. 1998. Modelling the seasonal cycle of dissolved organic carbon at Station E1 in the English Channel. *Estuarine, Coastal and Shelf Science* 46:93–109.
- Atkinson, L.P., D.W. Menzel, and K.A. Bush, eds. 1985. Oceanography of the Southeastern U.S. Continental Shelf, American Geophysical Union, Washington, DC, 156 pp.
- Bauer, J., and E. Druffel. 1998. Ocean margins as a significant source of organic matter to the open ocean. *Nature* 392:482–485.
- Behrenfeld, M.J., and P.G. Falkowski. 1997. Photosynthetic rates derived from satellite-based chlorophyll concentration. *Limnology and Oceanography* 42:1–20.
- Benner, R., J.D. Pakulski, M. McCarthy, J.I. Hedges, and P.G. Hatcher. 1992. Bulk chemical characteristics of dissolved organic matter in the ocean. *Science* 255:1,561–1,564.
- Biddanda, B., and R. Benner 1997. Carbon, nitrogen, and carbohydrate fluxes during the production of particulate and dissolved organic matter by marine phytoplankton. *Limnology and Oceanography* 42(3):506–518.
- Boyer, T.P., J.I. Antonov, H.E. Garcia, D.R. Johnson, R.A. Locarnini, A.V. Mishonov, M.T. Pitcher, O.K. Baranova, and I.V. Smolyar, 2006. World Ocean Database 2005. S. Levitus, ed., NOAA Atlas NESDIS 60, US Government Printing Office, Washington, DC, 190 pp., DVDs.

Carlson, C.A. 2002. Production and removal processes. Pp. 91–151 in *Biogeochemistry of Marine Dissolved Organic Matter*, D.A. Hansell and C.A. Carlson, eds, Academic Press, New York.

Carr, M.E., M.A.M. Friedrichs, M. Schmeltz, A.

Noguchi Aita, D. Antoine, K.R. Arrigo, I. Asanuma, O. Aumont, R. Barber, M. Behrenfeld, and others. 2006. A comparison of global estimates of marine primary production from ocean color. *Deep-Sea Research Part II* 53:741–770.

- Chassignet, E.P., H.E. Hurlbert, O.M. Smedstad, G.R. Halliwell, P.J. Hogan, A.J. Wallcraft, R. Baraille, and R. Bleck. 2007. The HYCOM (HYbrid Coordinate Ocean Model) data assimilative system. *Journal of Marine Systems* 65:60–83.
- Doney, S.C. 1999. Major challenges confronting marine biogeochemical modeling. *Global Biogeochemical Cycles* 13:705–714.
- Doney, S.C., I. Lima, K. Lindsay, J.K. Moore, S. Dutkiewicz, M.A.M Friedrichs, and R.J. Matear. 2001. Marine biogeochemical modeling: Recent advances and future challenges. *Oceanography* 14:93–107.
- Druffel, E., P. Williams, J. Bauer, and J. Ertel. 1992. Cycling of dissolved and particulate organic matter in the open ocean. *Journal of Geophysical Research* 97(C10):15,639–15,659.
- D'Sa, E.J., and R.L. Miller. 2003. Bio-optical properties in waters influenced by the Mississippi River during low flow conditions. *Remote Sensing of Environment* 84:538–549.
- Egbert, G.D., and S.Y. Erofeeva. 2002. Efficient inverse modeling of barotropic ocean tides. *Journal of Atmospheric and Oceanic Technology* 19:183–204.
- Evans, R.H., K.S. Baker, O.G. Brown, and R.C. Smith. 1986. Chronology of warm-core ring 82B. Journal of Geophysical Research 90:8,803–8,812.
- Fairall, C.W., E.F. Bradley, J.E. Hare, A.A. Grachev, and J.B. Edson. 2003. Bulk parameterization of air-sea fluxes: Updates and verification for the COARE algorithm. *Journal of Climate* 16:16,571–16,590.
- Fasham, M.J.R., H.W. Ducklow, and S.M. McKelvie. 1990. A nitrogen-based model of plankton dynamics in the oceanic mixed layer. *Journal of Marine Research* 48:591–639.
- Fennel, K., M. Losch, J. Schröter, and M. Wenzel. 2001. Testing a marine ecosystem model: Sensitivity analysis and parameter optimization. *Journal of Marine Systems* 28:45–63.
- Fennel, K., J. Wilkin, J. Levin, J. Moisan, J. O'Reilly, and D. Haidvogel. 2006. Nitrogen cycling in the Middle Atlantic Bight: Results from a threedimensional model and implications for the North Atlantic nitrogen budget. *Global Biogeochemical Cycles* 20: doi:10.1029/2005GB002456.

Flynn, K.J. 2005. Castles built on sand: Dysfunctionality in plankton models and the inadequacy of dialogue between biologists and modelers. *Journal of Plankton Research* 12:1,205–1,210.

- Friedrichs, M.A.M., R. Hood, and J. Wiggert. 2006. Ecosystem model complexity versus physical forcing: Quantification of their relative impact with assimilated Arabian Sea data. *Deep-Sea Research Part II* 53:576–600.
- Friedrichs, M.A.M, J.D. Dusenberry, L.A. Anderson, R. Armstrong, F. Chai, J.R. Christian, S.C. Doney, J. Dunne, M. Fujii, R. Hood, and others. 2007.

Assessment of skill and portability in regional marine biogeochemical models: Role of multiple planktonic groups. *Journal of Geophysical Research* 112(C08001): doi:10.1029/2006JC003852.

Garcia, H.E., and L.I. Gordon. 1992. Oxygen solubility in seawater: Better fitting equations. *Limnology and Oceanography* 37:1,307–1,312.

Gelinas, Y., J. Baldock, and J. Hedges. 2001. Organic carbon composition of marine sediments: Effect of oxygen exposure on oil generation potential. *Science* 294:145–148.

Haidvogel, D.B., H. Arango, W.P. Budgell, B.D. Cornuelle, E. Curchitser, E. Di Lorenzo, K. Fennel, W.R. Geyer, A.J. Hermann, L. Lanerolle, and others. In press. Regional ocean forecasting in terrainfollowing coordinates: Model formulation and skill assessment. Journal of Computational Physics.

Hansell, D.A., and C.A. Carlson. 1998. Deep-ocean gradients in the concentration of dissolved organic carbon. *Nature* 395:263–266.

Henrichs, S., and W. Reeburgh. 1987. Anaerobic mineralization of marine sediment organic matter: Rates and the role of anaerobic processes in the oceanic carbon economy. *Geomicrobiology Journal* 5(19):1–237.

Hofmann, E.E., and M.A.M. Friedrichs. 2001.
Biogeochemical data assimilation. Pp. 302–308 in Encyclopedia of Ocean Sciences, Vol. 1. J.H. Steele, S.A. Thorpe, and K.K. Turekian, eds, Academic Press, London, UK..

Hopkinson, C.S., and J.J. Vallino. 2005. Efficient export of carbon to the deep ocean through dissolved organic matter. *Nature* 433:142–145.

Ianson, D., and S.E. Allen 2002. A two-dimensional carbon and nitrogen flux model in a coastal upwelling region. *Global Biogeochemical Cycles* 16(1011): doi:10.1029/2001GB001451.

Johannessen, S.C., W.L. Miller, and J.J. Cullen. 2003. Calculation of UV attenuation and colored dissolved organic matter absorption spectra from measurements of ocean color. *Journal of Geophysical Research* 108(3301): doi:10.1029/2000JC000514.

Joliff, J., J.C. Kindle, I. Shulman, B. Penta, M.A.M. Friedrichs, R. Helber, and R.A. Arnone. In press. Summary diagrams for coupled hydrodynamicecosystem model skill assessment. *Journal of Marine Systems*, special issue on Skill Assessment for Coupled Biological/Physical Models of Marine Systems.

Lee, T.N., and L.P. Atkinson. 1983. Low frequency current and temperature variability from Gulf Stream frontal eddies and atmospheric forcing along the U.S. outer continental shelf. *Journal of Geophysical Research* 88:4,541–4,567.

Lozier, M.S., W.B. Owens, and R.G. Curry. 1995. The climatology of the North Atlantic. *Progress in Oceanography* 36:1–44.

Lozier, M.S., and G. Gawarkiewicz. 2001. Crossfrontal exchange in the Middle Atlantic Bight as evidenced by surface drifters. *Journal of Physical Oceanography* 31:2,498–2,510. Maritorena, S., D.A. Siegel, and A.R. Peterson. 2002. Optimization of a semianalytical ocean color model for global-scale applications. *Applied Optics* 41:2,705–2,714.

McClain, C.R., L.J. Pietrafesa, and J.A. Yoder. 1984. Observations of Gulf Stream-induced and winddriven upwelling in the Georgia Bight using ocean color and infrared images. *Journal of Geophysical Research* 89:3,705–3,723.

McClain, C.R., J.A. Yoder, L.P. Atkinson, J.O. Blanton, T.N. Lee, J.J. Singer, and F. Muller-Karger. 1988. Variability of surface pigment concentrations in the South Atlantic Bight. *Journal of Geophysical Research* 93:10,675–10,697.

NRC. 2005. Thinking Strategically: The Appropriate Use of Metrics for the Climate Change Science Program. The National Academies Press, Washington, DC, 150 pp.

Oke, P.R., J.S. Allen, R.N. Miller, G.D. Egbert, J.A. Austin, J.A. Barth, T.J. Boyd, P.M. Kosro, and M.D. Levine. 2002. A modeling study of the three-dimensional continental shelf circulation off Oregon. Part I: Model-data comparisons. *Journal of Physical Oceanography* 32:1,360–1,382.

O'Reilly, J.E., C. Zetlin, and D.A. Busch. 1987. Primary production. Pp. 221–233 in *Georges Bank*. R.H. Backus and D.W. Bourne, eds, MIT Press, Cambridge, MA.

Prézelin, B.B., E.E. Hofmann, M. Moline, and J.M. Klinck. 2004. Physical forcing of phytoplankton community structure and primary production in continental shelf waters of the western Antarctic Peninsula. *Journal of Marine Research* 62:419–460.

Ryan, J.P., J.A. Yoder, and D.W. Townsend. 2001. Influence of a Gulf Stream warm-core ring on water mass and chlorophyll distributions along the southern flank of Georges Bank. *Deep-Sea Research Part II* 48:159–178.

Shchepetkin, A.F., and J.C. McWilliams. 2005. The regional oceanic modeling system (ROMS): A split-explicit, free-surface, topography-following-coordinate ocean model. *Ocean Modelling* 9:347–404.

Seitzinger, S.P., and A.E. Giblin. 1996. Estimating denitrification in North Atlantic continental shelf sediments. *Biogeochemistry* 35:235–260.

Seitzinger, S.P., J.A. Harrison, E. Dumont, A.H.W. Beusen, and A.F. Bouwman. 2005. Sources and delivery of carbon, nitrogen and phosphorous to the coastal zone: An overview of global nutrient export from watersheds (NEWS) models and their application. Global Biogeochemical Cycles 19(4): GB4S01.

Signorini, S.R., C.R. McClain, A. Mannino, and S. Bailey. 2005. Report on ocean color and carbon study for the South Atlantic Bight and Chesapeake Bay regions. NASA/TM-2005-212787, August, Greenbelt, MD, 45 pp.

Soetaert, K., J.J. Middelburg, P.M.J. Herman, and K. Buis. 2000. On the coupling of benthic and pelagic biogeochemical models. *Earth-Science Review* 51:172–201. Søndergaard, M., P.J. le B. Williams, G. Cauwet, B. Riemann, C. Robinson, S. Terzic, E. Malcolm, S. Woodward, and J. Worm. 2000. Net accumulation and flux of dissolved organic carbon and dissolved organic nitrogen in marine plankton communities. *Limnology and Oceanography* 45(5):1,097–1,111.

Taylor, K.E. 2001. Summarizing multiple aspects of model performance in a single diagram. *Geophysical Research Letters* 106:7,183–7,192.

Thomas, C.J., N.E. Blair, M.J. Alperin, D.J. DeMaster, R.A. Jahnke, C.S. Martens, and L. Mayer. 2002. Organic carbon deposition on the North Carolina continental slope off Cape Hatteras (USA). *Deep-Sea Research Part II* 49:4,687–4,709.

Vlahos, P., R.F. Chen, and D.J. Repeta. 2002. Dissolved organic carbon in the Mid-Atlantic Bight. *Deep-Sea Research Part II* 49(20):4,369–4,385.

Walsh, J.J. 1972. Implications of a systems approach to oceanography. *Science* 176:969–975.

Wanninkhof, K. 1992. Relationship between wind speed and gas exchange over the ocean. Journal of Geophysical Research 97:7,373–7,382.

Warner J.C., C.R. Sherwood, R.P. Signell, C. Harris, and H.G. Arango. In press. Development of a three-dimensional, regional, coupled wave-, current-, and sediment-transport model. *Computers* and Geosciences.

Williams, P.J. le B. 1995. Evidence for the seasonal accumulation of carbon-rich dissolved organic material, its scale in comparison with changes in particulate material and the consequential effect on net C/N assimilation ratios. *Marine Chemistry* 51:17–29.

Wofsy, S.C., and R.C. Harriss. 2002. The North American Carbon Program (NACP). Report of the NACP Committee of the US Interagency Carbon Cycle Science Program. Washington, DC, US Global Change Research Program, 56 pp.

Yoder, J.A., 1985. Environmental control of phytoplankton production on the southeastern U.S. continental shelf. Pp. 93–103 in Oceanography of the Southeastern U.S. Continental Shelf. L.P. Atkinson, D.W. Menzel, and K.A. Bush, eds, American Geophysical Union, Washington, DC.

Yoder, J.A., L.P. Atkinson, S.S. Bishop, J.O. Blanton, T.N. Lee, and L.J. Pietrafesa. 1985. Phytoplankton dynamics within Gulf Stream intrusions on the southeastern United States continental shelf during summer 1981, *Continental Shelf Research* 4:611–635.

Yoder, J.A., C.R. McClain, J.O. Blanton, and L.-Y. Oey. 1987. Spatial scales in CZCS-chlorophyll imagery of the southeastern U.S. continental shelf. *Limnology and Oceanography* 32:929–941.

Yoder, J.A., J.E. O'Reilly, A.H. Barnard, T.S. Moore, and C.M. Ruhsam. 2001. Variability in coastal zone color scanner (CZCS) chlorophyll imagery of the ocean margin waters off the US east coast. *Continental Shelf Research* 21:1,191–1,218.

Zeebe, R., and D. Wolf-Gladrow. 2001. CO<sub>2</sub> in Seawater: Equilibrium, Kinetics, Isotopes. Elsevier, Amsterdam, 360 pp.