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Food for Thought

The Omega myth: what really drives lower calcification rates in an acidifying ocean

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The literature on ocean acidification (OA) contains a prevalent misconception that reduced organismal calcification rates in an acidifying ocean are driven by a reduction in carbonate ion (CO_3^{2-}) substrate availability (e.g. Omega or Ω). However, recent research in diverse organisms suggests that a reduction in seawater pH (i.e. increasing proton concentrations, $[H^+]$) is the most likely driver of reduced calcification rates in these organisms. OA leads to higher $[H^+]$ in seawater which alters the proton gradient between internal cellular reservoirs and external bulk seawater, making it difficult for organisms to maintain pH homeostasis. Biologically mediated calcification is a complex process, so it is unlikely that simple CO_3^{2-} substrate limitation is responsible for the observed decreases in calcification rates under OA conditions. Despite these inherent complexities, current predictions concerning the fate of calcifying organisms in an acidifying ocean have relied on the relationship between calcification rates and Ω . To more accurately predict how OA will affect the calcification of marine organisms, and consequently the global carbon cycle, we need to further elucidate the mechanisms driving observed decreases in calcification under acidified conditions.

Keywords: calcification, coral reef, ocean acidification, Omega, phytoplankton, saturation state.

Ocean acidification (OA) refers to the unprecedented reduction in seawater pH caused by anthropogenic CO₂ inputs (Hönisch *et al.*, 2012). OA is expected to reduce the ability of marine organisms such as corals, coccolithophores, foraminifera, and molluscs to secrete calcium carbonate (CaCO₃) skeletons, a process known as calcification (see Table 1) (Chan and Connolly, 2012; Kroeker *et al.*, 2013). As CO₂ dissolves into seawater, it lowers the pH and shifts the carbonate equilibria, decreasing the carbonate ion concentration ($[CO_3^{2-}]$). This lowers a chemical property of seawater known as the calcium carbonate saturation state or Ω . Seawater Ω is a function of CO_3^{2-} and calcium ion concentrations ($[Ca^{2+}]$) as follows

$$\Omega = \frac{[Ca^{2+}][CO_3^{2-}]}{K_{sp}},$$
(1)

where K_{sp} is the solubility product of a specific CaCO₃ mineral phase (e.g. aragonite or calcite) at a specified temperature, salinity, and

pressure (Zeebe and Wolf-Gladrow, 2001). Therefore, the thermodynamics of inorganic CaCO₃ precipitation and dissolution can largely be described by seawater Ω , with precipitation occurring at $\Omega > 1$ and dissolution at $\Omega < 1$ (Morse and Arvidson, 2002).

$$Ca^{2+} + CO_3^{2-} \leftrightarrow CaCO_3.$$
 (2)

From an OA perspective, Ω is mainly controlled by changing $[CO_3^{2-}]$, which is lowered as anthropogenic CO₂ dissolves into seawater, while $[Ca^{2+}]$ remains unaffected.

Having the thermodynamic principles of inorganic CaCO₃ precipitation in mind, early OA work investigated the calcification rates of organisms such as corals in seawater of varying Ω by manipulating bulk seawater [CO₃²⁻] and [Ca²⁺] (e.g. Gattuso *et al.*, 1998; Langdon *et al.*, 2000). Experimental evidence showed a positive correlation between calcification and Ω , which led to the idea that seawater [CO₃²⁻] could drive calcification rates. However, to determine

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Term	Definition
Calcification or gross calcification	Refers to the biologically controlled process of CaCO ₃ production, often occurs isolated from bulk seawater in media called the calcifying fluid
CaCO ₃ precipitation	Refers to the inorganic formation of CaCO ₃ minerals from a super saturated solution
CaCO ₃ dissolution	Refers to the inorganic dissolution of CaCO3 minerals in an under saturated solution, sometimes decalcification is used synonymously
Net calcification	The net effect of gross calcification and dissolution, usually refers to individual organisms
Net community calcification or net ecosystem calcification	The net effect of gross calcification and dissolution in an entire ecosystem

Table 1. Definition of terms referring to calcification and inorganic CaCO₃ precipitation/dissolution in the OA literature.

how OA affects calcification it becomes critical to differentiate between the inorganic precipitation and dissolution of CaCO₃ (which is thermodynamically constrained by seawater Ω) and biologically mediated calcification (see Table 1). In fact, it has been suggested that inorganic CaCO₃ dissolution may be more of a threat to marine calcifying organisms and calcareous ecosystems than decreasing calcification rates under OA (Andersson et al., 2009; Roleda et al., 2012; Eyre et al., 2014). However, many OA studies have not been able to isolate the effects of seawater Ω on organismal calcification rates in the absence of dissolution (i.e. gross calcification) because CaCO₃ is exposed to bulk seawater as either sediment or skeletal material. Therefore, any dissolution of exposed CaCO₃ could produce or enhance the observed relationships between Ω and net calcification. On the other hand, gross calcification is under biological control and mediated by organic tissue that separates the calcifying surface from overlying seawater. Therefore calcification occurs in a media (i.e. the calcifying fluid) that has significantly different $[CO_3^{2-}]$ than the bulk seawater. However, despite the complexities inherent to biological mediated calcification, much of the current OA literature presents the problem of reduced calcification under OA scenarios as an issue of simple CO_3^{2-} substrate availability [e.g. Equation (2)] (Hendriks *et al.*, 2015).

Recent work has demonstrated that corals can actively control carbonate chemistry at the site of calcification (Venn *et al.*, 2011; McCulloch *et al.*, 2012), which brings into question the mechanistic understanding of how external seawater Ω could influence organismal calcification rates. Currently, lines of research in different organisms and ecosystems are beginning to reach the same conclusion, that external seawater Ω (i.e. $[CO_3^{2-}]$) is not what drives changes in calcification rates. Here we show that these insights have been developed in two vastly different systems; scleractinian corals and open ocean phytoplankton known as coccolithophores.

The ability to modify carbonate chemistry at the site of calcification can produce internal conditions that are more thermodynamically favourable for inorganic CaCO₃ precipitation than in the surrounding seawater (i.e. a greater Ω). It has been shown that corals transport seawater to the site of calcification, however, once it is part of the calcifying fluid the chemistry of that seawater is actively modified (Gagnon et al., 2012). Coccolithophores passively regulate cytosolic pH through voltage-gated H⁺ channels; however, they actively regulate carbonate chemistry in their calcifying vesicles (Mackinder et al., 2010). Marine organisms have also been shown to modulate calcification through the use of organic molecules that can both stimulate or inhibit specific crystal lattice structures (Marsh, 1994). Therefore, in order for the external seawater chemistry to affect calcification rates, it must somehow affect that organism's ability to modulate its internal carbonate chemistry or produce organic compounds that mediate precipitation. Two lines of physiological evidence suggest that bulk seawater Ω does not control the Ω in the calcifying fluid, and thus, is not the major factor controlling organismal calcification rates.

1. While $\text{CO}_3^{2^-}$ is the substrate for inorganic CaCO₃ precipitation, no $\text{CO}_3^{2^-}$ transporter has been described in corals (Goiran *et al.*, 1996) or coccolithophores (Mackinder *et al.*, 2010). However, there is ample evidence that bicarbonate (HCO₃⁻) is actively transported into the calcifying fluid of corals (Goiran *et al.*, 1996; Moya *et al.*, 2008; Jury *et al.*, 2010) and coccolithophores (Berry *et al.*, 2002; Herfort *et al.*, 2002, Rost *et al.* 2003). Therefore, once HCO₃⁻ is transported into the calcifying fluid and $\text{CO}_3^{2^-}$ is combined with Ca²⁺ during calcification, H⁺ ions build-up (Figure 1);

$$Ca^{2+} + HCO_3^- \rightarrow H^+ + CaCO_3$$
 (3)

Since there is ~ 9 times the [HCO₃⁻] as [CO₃²⁻] in seawater, and OA increases [HCO₃⁻], it is unlikely that HCO₃⁻ substrate limitation under decreasing seawater pH is a problem for calcification. However, increasing seawater [H⁺] could be quite problematic.

2. As H⁺ ions build-up in the calcifying fluid the pH is lowered and the carbonate system shifts away from CO_3^{2-} , thus lowering Ω . Therefore, to maintain conditions favourable to inorganic CaCO₃ precipitation within the calcifying fluid, the organism must passively or actively remove H⁺ ions through membrane channels/transporters (Allemand *et al.*, 2011; Taylor *et al.*, 2012). As the oceans absorb CO₂ and seawater H⁺ concentrations increase, the electrochemical gradient between coral tissue and cytosolic fluids will decrease, making it more difficult to maintain high Ω in the calcifying fluid. This is the underlying concept behind the proton flux limitation model of calcification.

The conceptual framework of the proton flux limitation model has been elucidated in both corals (Jokiel, 2011a, b) and coccolithophores (Suffrian *et al.*, 2011; Bach *et al.*, 2013) (Figure 1). Once protons are transported into the tissue or cytosolic fluid from the calcifying fluid they must be removed to the water column to maintain intracellular pH, which becomes more energetically demanding as the proton concentration of seawater increases. If proton flux limitation at the boundary layer between the organism and seawater is the limiting factor for calcification, then increased energy production should lead to higher calcification rates by allowing more H⁺ ions to be actively pumped out of the calcifying fluid. This has been demonstrated in two distinct pathways in corals: (i) more light (and increases in photosynthesis) results in increased calcification rates (Marubini *et al.*, 2001; Al-Horani *et al.*, 2003) and (ii) feeding the coral with



Figure 1. A simplified schematic demonstrating the internal build-up of protons during the calcification process in corals and coccolithophores. Corals (left panel) must dissipate excess protons produced by calcification through a boundary layer and into the water column as proposed by Jokiel (2011b). Internally, corals most likely actively pump HCO_3^- ions into the calcifying fluid where protons build-up as $CaCO_3$ is precipitated. To maintain favourable conditions for precipitation in the calcifying fluid, corals likely actively pump $2H^+$ out and Ca^{+2} in using a Ca^{+2} -ATPase (Allemand *et al.*, 2011). To maintain the pH inside their tissue corals must remove protons, which becomes more energetically demanding when the gradient between the tissue and seawater $[H^+]$ is less pronounced due to ocean acidification. Suffrian *et al.* (2011) demonstrated that internal cellular pH (pH_i) in coccolithophores like *Emiliania huxleyi* (right panel) is directly affected by the surrounding seawater pH. This is most likely because *E. huxleyi* uses passive gated H⁺ channels to control cytosolic pH, which are forced to work against a less pronounced H⁺ gradient in an acidifying ocean. Black arrows represent fluxes between the organism and external seawater while white arrows represent fluxes occurring within the organism.

plankton enhances calcification rates (Ferrier-Pages *et al.*, 2003; Houlbrèque *et al.*, 2004; Towle *et al.*, 2015). Another hypothesis explaining light enhanced coral calcification is in direct agreement with the proton flux limitation hypothesis. Moya *et al.* (2006) suggested that increased pH in the gut of coral polyps during the day (Furla *et al.*, 2000; Al-Horani *et al.*, 2003) decreases the gradient against which H⁺ ions are pumped out of the calcifying fluid. It has also been suggested that the carbon concentrating mechanism (CCM) used by coral endosymbionts during daytime photosynthesis, which produces OH⁻ ions, can absorb excess H⁺ (Furla *et al.*, 2000). In coccolithophores, the cytosolic pH is regulated by H⁺ efflux via passive voltage-gated H⁺ channels (Suffrian *et al.* 2011). However, under ocean acidification scenarios, the electrochemical gradient becomes less and less favourable, most likely affecting pH and Omega inside the calcifying vesicle. Results from field and mesocosm studies of coral reef ecosystems are also consistent with the proton flux limitation model. Venti *et al.* (2014) recently demonstrated that light and temperature had much greater control than seawater Ω on changes in seasonal calcification rates of corals in the field. Also, a diel hysteretic pattern between external seawater Ω and calcification rates, with the highest rates of calcification occurring before the daily peak in bulk seawater Ω , has been observed in all levels of coral ecosystems from carbonate sediments (Cyronak *et al.*, 2013b), to corals and macroalgae (Jokiel *et al.*, 2014), and entire ecosystems (McMahon *et al.*, 2013; Shaw *et al.*, 2015). The hysteretic pattern observed in these studies is most likely due to the influence that benthic organisms have on seawater carbonate chemistry in coral reef ecosystems, with benthic production and calcification driving diel changes in seawater Ω , not the other way around. Importantly, these studies demonstrate the complexity in extrapolating the effects of OA based on short term natural changes in Ω . There is also a strong correlation between primary production and calcification in a range of studies (e.g. Gattuso *et al.*, 1999), indicating that photosynthesis is a dominant control on coral calcification.

Field and laboratory evidence across organisms and ecosystems indicates that calcification in an acidifying ocean is not controlled by bulk seawater Ω , but most likely inhibited by an increase in the gradient of protons between the calcifying fluid and external seawater. Therefore, a much better indicator of the influence of seawater chemistry on calcification may be the ratio of the substrate (dissolved inorganic carbon; DIC or $[HCO_2^-]$) to the waste product $([H^+])$ (Jokiel, 2011a, 2013). In fact, recent experimental evidence in mussels demonstrated that the seawater $[HCO_3^-]$ to $[H^+]$ ratio is important in controlling calcification rates (Thomsen et al., 2015), indicating that pH homeostasis may be important in controlling biologically mediated calcification in a diverse range of organisms. In contrast, a recent study suggested that bivalve larvae were particularly sensitive to Ω (Waldbusser *et al.*, 2015). However, Waldbusser *et al.* (2015) did not distinguish between the effects of Ω on gross calcification and dissolution, and their results may be due to the dissolution of CaCO₃ shells exposed to under saturated seawater. Furthermore, an additional explanation for the lack of correlative power between changes in calcification rates and pH (instead of Ω) in some studies with un-coupled carbonate chemistry manipulations, such as in Langdon et al. (2000) or Waldbusser et al. (2015), is that increasing $[HCO_3^-]$ could partially compensate for unfavourable pH levels.

It is also important to remember that calcification is influenced by many other factors such as light availability and temperature, which may be more important in driving future changes in calcification rates (McNeil *et al.*, 2004). The coupling of seawater pH and Ω under ocean acidification scenarios (i.e. inputs of CO₂) can lead to positive correlations between calcification and both pH and Ω . However, pH and Ω can become decoupled or negatively correlated on geological time scales due to land-based weathering processes (Hönisch *et al.*, 2012), and on more modern timescales due to opposite effects of temperature on pH and Ω (Figure 2) (Gattuso *et al.*, 1999). While this decoupling is not likely on the timescale of modern OA in the open ocean, it could occur in some coastal areas with pronounced diel temperature variability.



Figure 2. Correlation of pH and aragsonite saturation state (Ω) when temperature is increased from 20 to 30°C. pH and Ω were calculated at constant TA and DIC concentrations of 2300 and 2000 μ mol kg⁻¹, respectively, while temperature was varied by 1°C from 20 to 30°C. It is important to note that global warming will not drive the uncoupling of pH and Ω due to ongoing CO₂ equilibration between the atmosphere and ocean.

Current predictions of the fate of coral reefs in an acidifying ocean which are reliant on the relationship between net ecosystem calcification and Ω (e.g. Figure 6 in Shamberger *et al.*, 2011 could be based on a basic misconception about the factors driving changes in coral physiology. With this in mind, it becomes vitally important to grasp the correct mechanistic understanding of how increasing CO₂ inhibits the calcification of marine organisms. We are not the first to stress this point. A recent perspective article noted a disconnect between the more recent OA literature and older studies on calcification physiology (Roleda et al., 2012). Other recent work is beginning to highlight the sensitivity of CaCO₃ dissolution to OA, which may pose a more serious threat to coral reef ecosystems than changes in calcification (Andersson et al., 2009; Cyronak et al., 2013a; Eyre et al., 2014). Multiple theories exist for the influence of OA on marine calcifiers (Allemand et al., 2011). However, the prevailing notion in the OA literature that calcification is inhibited through a reduction in seawater $[CO_3^{2-}]$, and thus Ω , is most likely incorrect. Rather, as outlined above, it is most likely the decrease in seawater pH and associated problems of pH homeostasis within organisms that governs changes in calcification rates under OA conditions.

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References

- Al-Horani, F. A., Al-Moghrabi, S. M., and de Beer, D. 2003. The mechanism of calcification and its relation to photosynthesis and respiration in the scleractinian coral *Galaxea fascicularis*. Marine Biology, 142: 419–426.
- Allemand, D., Tambutté, É., Zoccola, D., and Tambutté, S. 2011. Coral calcification, cells to reefs. *In* Coral Reefs: An Ecosystem in Transition, pp. 119–150. Ed. by Z. Dubinsky, and N. Stambler. Springer, The Netherlands.
- Andersson, A. J., Kuffner, I. B., Mackenzie, F. T., Jokiel, P. L., Rodgers, K. S., and Tan, A. 2009. Net Loss of CaCO₃ from a subtropical calcifying community due to seawater acidification: mesocosm-scale experimental evidence. Biogeosciences, 6: 1811–1823.
- Bach, L. T., Mackinder, L. C. M., Schulz, K. G., Wheeler, G., Schroeder, D. C., Brownlee, C., and Riebesell, U. 2013. Dissecting the impact of CO2 and pH on the mechanisms of photosynthesis and calcification in the coccolithophore Emiliania huxleyi. New Phytologist, 199: 121–134.
- Berry, L., Taylor, A. R., Lucken, U., Ryan, K. P., and Brownlee, C. 2002. Calcification and inorganic carbon acquisition in coccolithophores. Functional Plant Biology, 29: 289–299.
- Chan, N. C. S., and Connolly, S. R. 2012. Sensitivity of coral calcification to ocean acidification: a meta-analysis. Global Change Biology, 19: 282–290.
- Cyronak, T., Santos, I., and Eyre, B. 2013a. Permeable coral reef sediment dissolution driven by elevated *p*CO₂ and pore water advection. Geophysical Research Letters, 40: 4876–4881.
- Cyronak, T., Santos, I., McMahon, A., and Eyre, B. D. 2013b. Carbon cycling hysteresis in permeable carbonate sands over a diel cycle: implications for ocean acidification. Limnology and Oceanography, 58: 131–143.
- Eyre, B. D., Andersson, A. J., and Cyronak, T. 2014. Benthic coral reef calcium carbonate dissolution in an acidifying ocean. Nature Clim. Change, 4: 969–976.

- Ferrier-Pages, C., Witting, J., Tambutté, E., and Sebens, K. 2003. Effect of natural zooplankton feeding on the tissue and skeletal growth of the scleractinian coral Stylophora pistillata. Coral Reefs, 22: 229–240.
- Furla, P., Allemand, D., and Orsenigo, M. N. 2000. Involvement of H(+)-ATPase and carbonic anhydrase in inorganic carbon uptake for endosymbiont photosynthesis. American Journal of Physiology: Regulatory, Integrative and Comparative Physiology, 278: R870–R881.
- Gagnon, A. C., Adkins, J. F., and Erez, J. 2012. Seawater transport during coral biomineralization. Earth and Planetary Science Letters, 329– 330: 150–161.
- Gattuso, J -P., Allemand, D., and Frankignoulle, M. 1999. Photosynthesis and calcification at cellular, organismal and community levels in coral reefs: A review on interactions and control by carbonate chemistry. American Zoologist, 39: 160–183.
- Gattuso, J. P., Frankignoulle, M., Bourge, I., Romaine, S., and Buddemeier, R. W. 1998. Effect of calcium carbonate saturation of seawater on coral calcification. Global and Planetary Change, 18: 37–46.
- Goiran, C., Al-Moghrabi, S., Allemand, D., and Jaubert, J. 1996. Inorganic carbon uptake for photosynthesis by the symbiotic coral/dinoflagellate association I. Photosynthetic performances of symbionts and dependence on sea water bicarbonate. Journal of Experimental Marine Biology and Ecology, 199: 207–225.
- Hendriks, I. E., Duarte, C. M., Olsen, Y. S., Steckbauer, A., Ramajo, L., Moore, T. S., Trotter, J. A., *et al.* 2015. Biological mechanisms supporting adaptation to ocean acidification in coastal ecosystems. Estuarine, Coastal and Shelf Science, 152: A1–A8.
- Herfort, L., Thake, B., and Roberts, J. 2002. Acquisition and use of bicarbonate by Emiliania huxleyi. New Phytologist, 156: 427–436.
- Hönisch, B., Ridgwell, A., Schmidt, D. N., Thomas, E., Gibbs, S. J., Sluijs, A., Zeebe, R., *et al.* 2012. The geological record of ocean acidification. Science, 335: 1058–1063.
- Houlbrèque, F., Tambutté, E., Allemand, D., and Ferrier-Pagès, C. 2004. Interactions between zooplankton feeding, photosynthesis and skeletal growth in the scleractinian coral Stylophora pistillata. Journal of Experimental Biology, 207: 1461–1469.
- Jokiel, P. L. 2013. Coral reef calcification: carbonate, bicarbonate and proton flux under conditions of increasing ocean acidification. Proceedings of the Royal Society of London B, 20130031. [doi:10.1098/rspb.2013.0031].
- Jokiel, P. L. 2011a. Ocean acidification and control of reef coral calcification by boundary layer limitation of proton flux. Bulletin of Marine Science, 87: 639–657.
- Jokiel, P. L. 2011b. The reef coral two compartment proton flux model: A new approach relating tissue-level physiological processes to gross corallum morphology. Journal of Experimental Marine Biology and Ecology, 409: 1–12.
- Jokiel, P. L., Jury, C. P., and Ku'ulei, S. R. 2014. Coral-algae metabolism and diurnal changes in the CO2-carbonate system of bulk sea water. PeerJ, 2: e378.
- Jury, C. P., Whitehead, R. F., and Szmant, A. M. 2010. Effects of variations in carbonate chemistry on the calcification rates of *Madracis auretenra* (= *Madracis mirabilis* sensu Wells, 1973): bicarbonate concentrations best predict calcification rates. Global Change Biology, 16: 1632–1644.
- Kroeker, K. J., Kordas, R. L., Crim, R., Hendriks, I. E., Ramajo, L., Singh, G. S., Duarte, C. M., *et al.* 2013. Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. Global Change Biology, 19: 1884–1896.
- Langdon, C., Takahashi, T., Sweeney, C., Chipman, D., Goddard, J., Marubini, F., Aceves, H., *et al.* 2000. Effect of calcium carbonate saturation state on the calcification rate of an experimental coral reef. Global Biogeochemical Cycles, 14: 639–654.
- Mackinder, L., Wheeler, G., Schroeder, D., Riebesell, U., and Brownlee, C. 2010. Molecular Mechanisms Underlying Calcification in Coccolithophores. Geomicrobiology Journal, 27: 585–595.

- Marsh, M. E. 1994. Polyanion-mediated mineralization—assembly and reorganization of acidic polysaccharides in the Golgi system of a coccolithophorid alga during mineral deposition. Protoplasma, 177: 108–122.
- Marubini, F., Barnett, H., Langdon, C., and Atkinson, M. 2001. Dependence of calcification on light and carbonate ion concentration for the hermatypic coral Porites compressa. Marine Ecology Progress Series, 220: 153–162.
- McCulloch, M., Falter, J., Trotter, J., and Montagna, P. 2012. Coral resilience to ocean acidification and global warming through pH up-regulation. Nature Climate Change, 2: 623–627.
- McMahon, A., Santos, I. R., Cyronak, T., and Eyre, B. D. 2013. Hysteresis between coral reef calcification and the seawater aragonite saturation state. Geophysical Research Letters, 40: 4675–4679.
- McNeil, B. I., Matear, R. J., and Barnes, D. J. 2004. Coral reef calcification and climate change: The effect of ocean warming. Geophysical Research Letters, 31: L22309.
- Morse, J. W., and Arvidson, R. S. 2002. The dissolution kinetics of major sedimentary carbonate minerals. Earth-Science Reviews, 58: 51–84.
- Moya, A., Tambutté, S., Bertucci, A., Tambutté, E., Lotto, S., Vullo, D., Supuran, C. T., *et al.* 2008. Carbonic Anhydrase in the Scleractinian Coral *Stylophora pistillata*. Journal of Biological Chemistry, 283: 25475–25484.
- Moya, A., Tambutte, S., Tambutte, E., Zoccola, D., Caminiti, N., and Allemand, D. 2006. Study of calcification during a daily cycle of the coral Stylophora pistillata: implications for 'light-enhanced calcification'. Journal of Experimental Biology, 209: 3413–3419.
- Roleda, M. Y., Boyd, P. W., and Hurd, C. L. 2012. Before ocean acidification: Calcifier chemistry lessons. Journal of Phycology, 48: 840–843.
- Rost, B., Riebesell, U., Burkhardt, S., and Sültemeyer, D. 2003. Carbon acquisition of bloom-forming marine phytoplankton. Limnology and Oceanography, 48: 55–67.
- Shamberger, K. E. F., Feely, R. A., Sabine, C. L., Atkinson, M. J., DeCarlo, E. H., Mackenzie, F. T., Drupp, P. S., *et al.* 2011. Calcification and organic production on a Hawaiian coral reef. Marine Chemistry, 127: 64–75.
- Shaw, E. C., Phinn, S. R., Tilbrook, B., and Steven, A. 2015. Natural in situ relationships suggest coral reef calcium carbonate production will decline with ocean acidification. Limnology and Oceanography. Doi:10.1002/lno.10048.
- Suffrian, K., Schulz, K. G., Gutowska, M., Riebesell, U., and Bleich, M. 2011. Cellular pH measurements in Emiliania huxleyi reveal pronounced membrane proton permeability. New Phytologist, 190: 595–608.
- Taylor, A. R., Brownlee, C., and Wheeler, G. L. 2012. Proton channels in algae: reasons to be excited. Trends in Plant Science, 17: 675–684.
- Thomsen, J., Haynert, K., Wegner, K. M., and Melzner, F. 2015. Impact of seawater carbonate chemistry on the calcification of marine bivalves. Biogeosciences Discussion, 12: 1543–1571.
- Towle, E. K., Enochs, I. C., and Langdon, C. 2015. Threatened Caribbean Coral Is Able to Mitigate the Adverse Effects of Ocean Acidification on Calcification by Increasing Feeding Rate. PLoS ONE, 10: e0123394.
- Venn, A., Tambutté, E., Holcomb, M., Allemand, D., and Tambutté, S. 2011. Live tissue imaging shows reef corals elevate ph under their calcifying tissue relative to seawater. PLoS ONE, 6: e20013.
- Venti, A., Andersson, A., and Langdon, C. 2014. Multiple driving factors explain spatial and temporal variability in coral calcification rates on the Bermuda platform. Coral Reefs, 33: 979–997.
- Waldbusser, G. G., Hales, B., Langdon, C. J., Haley, B. A., Schrader, P., Brunner, E. L., Gray, M. W., *et al.* 2015. Saturation-state sensitivity of marine bivalve larvae to ocean acidification. Nature Climate Change, 5: 273–280.
- Zeebe, R. E., and Wolf-Gladrow, D. 2001. CO₂ in Seawater: Equilibrium, Kinetics, Isotopes. Elsevier Oceanography Series, 65, Amsterdam.